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(11) Publication number:

0 139 383
A1

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 84305571.6

(51) Int. Cl. 4: C 12 N 15/00

(22) Date of filing: 16.08.84

C 12 P 21/00, C 12 N 9/99
A 61 K 37/02
//C12R1/85

(30) Priority: 16.08.83 US 523685

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(43) Date of publication of application:
02.05.85 Bulletin 85/18

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(54) Designated Contracting States:
AT BE CH DE FR GB IT LI LU NL SE

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(54) Method for expressing foreign genes in *schizosaccharomyces pombe* and the use in therapeutic formulations of the products, DNA constructs and transformant strains of *schizosaccharomyces pombe* usable in such method and their preparation.

(57) A method is provided for producing heterologous polypeptides in yeast *Schizosaccharomyces pombe*. DNA constructs are provided for use in this method comprising a segment coding for an *S. pombe* promoter. A particular construct is provided comprising the *S. pombe* alcohol dehydrogenase promoter and human alpha-1-antitrypsin gene, which is used to transform *S. pombe* to produce a substantially unglycosylated protein having activity of human alpha-1-antitrypsin.

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METHOD FOR EXPRESSING FOREIGN GENES IN SCHIZOSACCHAROMYCES POMBE AND THE USE IN THERAPEUTIC FORMULATIONS OF THE PRODUCTS, DNA CONSTRUCTS AND TRANSFORMANT STRAINS OF SACCHAROMYCES POMBE USABLE IN SUCH METHOD AND THEIR PREPARATION

5 The present invention is directed to methods for expressing heterologous polypeptides in the fission yeast Schizosaccharomyces pombe, to DNA vectors useful therein, and to protein products expressed thereby.

In the field of genetic engineering where the object is normally to obtain high expression of a polypeptide product, usually a mammalian protein, in a micro-
10 organism, the choice of the host organism may be critical. Some work has been done in the eucaryotic organism Saccharomyces cerevisiae (Baker's yeast). However, whether other eucaryotic microorganisms may achieve similar or even higher levels of expression of hetero-
15 logous proteins was not heretofore known. The fission yeast Schizosaccharomyces pombe is only distantly related to S. cerevisiae and the two genera differ in many aspects of biochemistry and physiology. Such differences, for example, may be reflected in possible
20 different levels and/or specificities of proteases in the different species which may degrade heterologous proteins. Also, the yeast genera may differ in enzymes which modify heterologous proteins, which may involve differing specific cleavage sites, sulphhydryl bridges, glycosylation, amino-terminal modifications and the like. Depending upon the particular heterologous

protein which one may express, such specific modifications may be desirable or undesirable. Furthermore, different yeast genera may contain different proteins which in some cases may be more difficult than others to 5 purify away from the desired heterologous protein. Also, at certain levels the presence of heterologous proteins may have a detrimental effect on cell viability, so it may be desirable to determine which yeast genera may be most tolerant to the particular desired 10 protein. The secretion processes may differ among yeast genera, therefore one may be able to take advantage of such properties, for example, a particular heterologous protein may be secreted at a higher level in one genus versus another, or undesirable proteases and protein 15 modifying enzymes may be secreted at a lower level while the heterologous protein is secreted at a higher level in certain genera. Finally, yeast genera may differ in their growth rates or yields in different media at different temperatures and pressures, thereby affecting 20 time and cost of production. It is therefore desirable to provide a method of producing foreign proteins in different eucaryotic organisms to take advantage of such differences in characteristics. In most cases, such differences in characteristics may not be determined a 25 priori.

It is therefore an aspect of the present invention to provide methods for producing heterologous proteins in the fission yeast Schizosaccharomyces pombe.

It is a further aspect of the present invention to 30 provide DNA vectors which are useful for expressing heterologous proteins in S. pombe.

It is another aspect of the present invention to provide DNA expression vectors comprising S. pombe alcohol dehydrogenase promoter.

5 It is a further aspect of the present invention to provide substantially pure unglycosylated human alpha-1 antitrypsin produced by Schizosaccharomyces pombe.

In the accompanying FIGURES:

FIG. 1 is the cDNA sequence of the gene coding for the predominant form of human alpha-1 antitrypsin.

10 FIG. 2 illustrates a partial sequence of pUC13 including the linker.

FIG. 3 illustrates a method of preparation of vector HAT 4.

15 FIG. 4 illustrates the restriction map of vector C1/1, a vector utilized in preparation of HAT 4.

FIG. 5 illustrates the restriction map of the vector pUC α -1 + FG1, an intermediate vector used in the preparation of HAT 4.

FIG. 6 illustrates the restriction map of HAT 4.

20 FIG. 7 illustrates a method of preparing vector pR103-5, an expression vector for producing alpha-1 antitrypsin in S. pombe.

FIG. 8 illustrates the restriction map of pR103-5.

The present invention is directed to a method for producing a heterologous protein in yeast genus S. pombe comprising the step of growing a culture of the yeast transformed by a DNA vector containing a segment coding 5 for an S. pombe promoter, whereby expression of the protein is under regulation of the promoter. DNA constructs useful for making such vectors are also provided. In particular, a vector is constructed comprising the S. pombe alcohol dehydrogenase promoter 10 and the structural gene for human alpha-1 antitrypsin which is transformed into S. pombe. Growth of the transformant produces a substantially unglycosylated protein having the activity of mammalian alpha-1 anti-trypsin. This protein may be isolated in a substantial- 15 ly pure form.

In order to express heterologous proteins in S. pombe, it is desirable to have the expression under control of a promoter which is indigenous to S. pombe. Such a promoter may be the approximately 700 base pair 20 Sph I-Eco RI fragment comprising the S. pombe ADH promoter which may be isolated from plasmid pADHpol, a plasmid which is disclosed by Russell et al., J.Biol.Chem., 258, 143-149 (1983), the disclosure of which is incorporated by reference herein in its 25 entirety. Other S. pombe promoters may include the glycolytic pathway promoters, such as, triose phosphate isomerase, phosphofructo kinase, glyceraldehyde-3-phosphate dehydrogenase. The heterologous protein selected for expression is alpha-1 antitrypsin, a 30 protease inhibitor present in mammalian blood whose apparently major physiological function is to inhibit elastase, a potent protease which hydrolyzes structural proteins. Alpha-1 antitrypsin may also inhibit other serine proteases. A low level of alpha-1 antitrypsin in

the blood may be associated with chronic obstructive pulmonary emphysema and infantile liver cirrhosis. Therefore, production of proteins having alpha-1 anti-trypsin activity may be useful for treating alpha-1 antitrypsin deficiency and to examine the mechanism of the acute-phase response which occurs under inflammatory conditions in the body whereby the concentration of alpha-1 antitrypsin is substantially increased.

5 The gene for the predominant form of human alpha-1 antitrypsin is shown in FIG. 1. A DNA fragment comprising such gene which may be utilized may be derived from the plasmid HAT 4. The construction of HAT 4 is illustrated generally in FIG. 3.

10 To prepare HAT 4, the intermediate vector C1/1 is utilized. Referring to FIG. 3, the S. cerevisiae TPI promoter, described by Alber and Kawasaki, J. Molec. Appl. Genet., 1, 419-434 (1982) is digested with Bal31 exonuclease from the KpnI restriction site within the TPI coding region. The resultant blunt end is converted to 15 a Eco RI sticky end by the addition of a 5' blunt end Eco RI adaptor. A synthetic oligonucleotide linker having the sequence AATTCATGGAGGATCC is ligated to the Eco RI sticky end, then digested with Bam HI. The resultant fragment comprising the TPI promoter may be 20 inserted into the Bam HI site of the vector C1/1. The restriction map of C1/1 is shown in FIG. 4. The C1/1 plasmid was constructed from plasmid pJDB248 (Beggs, Nature, 275, 104-109 (1978)) by removing the pMB9 25 sequences from pJDB248 by a partial digestion with Eco RI. Eco RI digested pBR322 was then inserted to give C1/1. It will be noted that the linker sequence, 30 in addition to providing Eco RI and Bam HI restriction sites, also supplies an ATG initiation codon and a GAG

codon, which codes for glutamic acid, the first amino acid of naturally occurring human alpha-1 antitrypsin. Therefore, these codons are "in frame" when the rest of the alpha-1 antitrypsin structural gene is inserted at 5 the Bam HI site.

The gene coding for the predominant form of human alpha-1 antitrypsin (shown in FIG. 1) may be isolated from a human cDNA library using the baboon sequence (Kurachi et al., PNAS USA, 78, 6826-6830 (1981); and 10 Chandra et al., Biochem.Biophys.Res.Com., 103, 751-758 (1981)) as a DNA hybridization probe. This gene may then be inserted as a PstI fragment, as shown in FIG. 3 into vector pUC13 (prepared as described by Vieira et al., Gene, 19, 259-268 (1982) for vectors pUC8 and pUC9, 15 but containing the multiple restriction site shown in FIG. 2 at start of the lacZ gene) at the single PstI site in pUC13. In the resultant construct the anti- trypsin gene was followed by an XbaI site and an Eco RI site in the multiple cloning sequence. Therefore, 20 between these two sites was inserted the yeast TPI terminator as a 700 base pair XbaI-Eco RI fragment from pTPIC10. The plasmid pTPIC10 is described by Alber and Kawasaki, J.Molec.App.Gene., 1, 419-434 (1982). The resultant plasmid shown in FIG. 3 is pUC α -1 + FG1 which 25 contains the human alpha-1 antitrypsin with a yeast transcription terminator. A detailed structure of pUC α -1 + FG1 is shown in FIG. 5.

Referring to FIG. 3 an Eco RI, Bam HI synthetic DNA adaptor is then added to the Eco RI site of pUC α -1 + FG1 30 in order to create a Bam HI site on the 3' end of the yeast terminator. By using this adaptor the human alpha-1 antitrypsin yeast terminator sequence may be removed by cutting with Bam HI to liberate a fragment of

approximately 2100 base pairs. This Bam HI fragment is then inserted into the modified C1/l plasmid containing the TPI promoter with Bam HI adaptor. The resultant plasmid HAT 4 contains the TPI promoter, ATGGAG adaptor, 5 human alpha-1 antitrypsin gene and TPI terminator. A detailed structure of HAT 4 is shown in FIG. 6. Referring to FIG. 7, HAT 4 was digested with Eco RI and Hind III and the approximately 1400 base pair Hind III-Eco RI fragment was isolated. This fragment was ligated to the 10 Eco RI-Sph I 700 base pair fragment containing the S. pombe ADH promoter discussed above. The resultant Hind III-Sph I linear segment was then inserted into Sph I and Hind III digested shuttle vector pCAS. Plasmid pCAS is a derivative of CV13 (also known as YEpl3; 15 Broach et al., Gene, 8, 121-123 (1979)) in which the single SstI site has been destroyed by cutting with SstI, blunting the ends with T4 DNA polymerase and dNTP's, and religating the linear molecule. The final construct shown in FIGURE 7 containing the S. pombe ADH 20 promoter, alpha-1 antitrypsin coding sequence, and TPI terminator is known as pR103-5. A detailed structure of pR103-5 is shown in FIG. 8. This plasmid also contains the S. cerevisiae LEU2 gene which complements the leu1-32 mutation of S. pombe.

25 The plasmid pR103-5 may be used to transform S. pombe strains by conventional techniques. Growth of the transformed cells will result in expression of polypeptides exhibiting the protease inhibition activity of human alpha-1 antitrypsin. These proteins may be useful 30 for treatment of genetic alpha-1 antitrypsin deficiency and other disease states related to inadequate levels of the alpha-1 antitrypsin. Thus, conditions such as emphysema and other lung disorders related to progressive digestion of lung sacs may be treated, such as,

chronic obstructive pulmonary disease or adult respiratory distress syndrome. Non-genetically related emphysema may be also be treated, such as, emphysema resulting from heavy smoking. Conditions not necessarily 5 confined to the lungs may also be treated, such as, cystic fibrosis and arthritis. For a review of alpha-1 antitrypsin in deficiency see Gadek, J. E., and R. Crystl, "Alpha-1 Antitrypsin Deficiency", The Metabolic Basis of Inherited Disease, J. B. Stanbury, J. B. 10 Wyngaarden and D. S. Fredrickson, McGraw-Hill, New York, pages 1450-1467 (1982).

The proteins according to the present invention may be admixed with conventional pharmaceutical carriers. Preferably the proteins may be administered intravenously or by inhalation. While effective dosages may 15 vary according to the severity of the condition and weight of the subject, dosages in the range of 0.5 to 10.0 gm/wk of a protein introduced intraveneously may, in many cases, be effective. Lower dosages may be 20 effective if the method of administration is by inhalation. Oral administration may also be effective provided the alpha-1 antitrypsin is protected in capsules or coded carriers from premature degradation in the digestive tract. The following examples set forth 25 specific embodiments according to the present invention, but the invention is not intended to be limited thereto.

EXAMPLE 1

EXPRESSION OF PROTEINS HAVING ALPHA-1 ANTITRYPSIN ACTIVITY FROM S. POMBE.

30 The pR103-5 expression vector is used to transform S. pombe strain PR118-4 (h-, leu1-32), ATCC Deposit

No. 20680. The strain is obtained by crossing strain PR131 (h+, leu1-32), described by Kohli et al., Genetics, 87, 471-489 (1977), and strain PRP17 (h-, cdc10-129), described by Nurse et al., Molecular and General Genetics, 146, 167-168 (1976), and isolating clones which give consistent colony size. The transformed cells may be grown under selective conditions described by Russell et al., Nature, 301, 167-169 (1983). RNA analysis using Northern blots (Thomas, PNAS USA, 77, 5201-5205 (1980)) and a ³²P-labeled Eco RI-Hind III alpha-1 antitrypsin-TPI terminator probe indicates that high level of transcription of alpha-1 antitrypsin cDNA is directed by the ADH promoter. Cell extracts from the log phase cultures grown under selective conditions are analyzed by ELISA assay and show 0.85%-0.95% of total soluble protein to be protein exhibiting alpha-1 antitrypsin activity.

EXAMPLE 2

CHARACTERIZATION OF PROTEINS FROM S. POMBE EXHIBITING 20 ALPHA-1 ANTITRYPsin ACTIVITY.

Alpha-1 antitrypsin proteins from the S. pombe transformants above in Example 1 are further characterized by a purification on amino-adsorption column and electrophoresis on a polyacrylamide/SDS gel. The column is prepared by covalently attaching affinity purified antibodies to human alpha-1 antitrypsin to CNBr-activated Sepharose according to the method of Cuatrecasas, J.Biochem., 245, 3059 (1970). Disrupted cells are extracted with phosphate buffered saline pH 7.2 containing 0.5m NaCl and applied to the column. The column is eluted with 3m NaSCN and the recovered material is analyzed by electrophoresis on a polyacrylamide

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gel in the presence of sodium dodecyl sulfate. The alpha-1 antitrypsin proteins which are produced in S. pombe have molecular weights in the range of 42-43 kdal. Naturally occurring human alpha-1 antitrypsin has a 5 molecular weight of approximately 54 kdal., having a carbohydrate composition of approximately 16% by weight. See Hodges et al., J.Biochem., 254, 8208-8212 (1979). The proteins having alpha-1 antitrypsin activity produced by S. pombe will thus appear to be 10 unglycosylated or substantially unglycosylated.

EXAMPLE 3

ELISA Assay for AT

To assay for AT, 200 microliters of goat anti-alpha-1- 15 antitrypsin (10 micrograms/ml in 0.1M Na_2CO_3 pH 9.8) that has been affinity purified is placed in each well of a 96 well microtiter culture plate at 37°C for two to four hours. This solution is removed and 200 microliters of phosphate buffered saline (PBS) pH 7.2 containing 1% BSA and .05% Tween 20 is added to each 20 well at 37°C for two hours.

Test samples, including standards, are then added to each well and AT, if present, binds to the specific antibody. The wells are rinsed with PBS after 15-30 25 minutes of incubation. 200 microliters of rabbit anti-alpha-1-AT is added to each well in the presence of 1% BSA in PBS. After 15 minutes incubation, the solution is removed and the wells are rinsed with PBS.

A sample of 200 microliters of goat anti rabbit IgG coupled to alkaline phosphatase in 1% BSA and PBS are 30 added to each well and incubated for 15 minutes. After

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rinsing the wells, 200 microliters of a substrate
solution was added which contains 30 milligrams of
dinitrophenyl phosphate dissolved in 50 ml of 0.1 m
diethanolamine pH 9.6. A yellow color is allowed to
5 develop (15-60 minutes) and intensity is recorded at 405
nm on a Titertek Multiscan detector. The yellow color
is proportional to the amount of AT in the test solu-
tion.

CLAIMS:

1. A method of producing a heterologous polypeptide in yeast Schizosaccharomyces pombe comprising the step of growing a culture of said yeast transformed by a DNA 5 vector, said vector comprising a segment coding for an S. pombe promotor, wherein expression of said polypeptide is under regulation of said promotor.
2. A method as claimed in claim 1, wherein said promotor comprises the alcohol dehydrogenase promotor. 10
3. A method as claimed in claim 1 or claim 2, wherein said polypeptide possesses the protease inhibition activity of mammalian alpha-1-antitrypsin.
4. A method as claimed in claim 3, wherein said polypeptide has the amino acid sequence of a form, e.g. 15 the predominant form, of mammalian alpha-1-antitrypsin.
5. A method as claimed in claim 1, wherein said vector comprises plasmid pR103-5.
6. A method as claimed in any one of claims 1 to 5, wherein said polypeptide is substantially unglycosylated. 20
7. A method as claimed in any one of claims 1 to 6, wherein said polypeptide is substantially pure.
8. A DNA construct comprising a promotor of S. pombe and a gene for a heterologous protein positioned such that expression of said protein in S. pombe is under regulation 25 of said promotor.
9. A construct as claimed in claim 8 and further defined by the specific feature of any one of claims 2 to 4.
10. A DNA construct as claimed in claim 8 comprising the DNA sequence of pR103-5. 30
11. A method of constructing a DNA vector capable in S. pombe of expressing a heterologous polypeptide, e.g. mammalian alpha-1-antitrypsin, comprising ligating DNA segments comprising respectively a structural gene 35 encoding said polypeptide and a promotor indigenous to S. pombe.

12. A transformant strain of S. pombe containing a DNA construct as defined in any one of claims 8 to 10.

13. A method of forming a transformant strain of S. pombe having the capability of producing a heterologous 5 polypeptide, e.g. a polypeptide having the protease inhibition activity of mammalian alpha-1-antitrypsin, which method comprises introducing a DNA transfer vector comprising a construct as defined in any one of claims 8 to 10 into S. pombe to transform the same thereby 10 altering its genotype to encode in expressible format for said polypeptide.

14. A therapeutic formulation for treating disease states related to inadequate amounts of alpha-1-antitrypsin in a mammal comprising a polypeptide which 15 has been produced by a method as claimed in any one of claims 3 to 5 or in claim 6 or claim 7 when dependent directly or indirectly upon any one of claims 3 to 5 formulated for such treatment, e.g. for administration by inhalation, for oral administration or for intravenous 20 administration.

15. A method of producing a therapeutic formulation for treating disease states related to inadequate amounts of alpha-1-antitrypsin in a mammal comprising formulating for such treatment a polypeptide which has been produced 25 by a method as claimed in any one of claims 3 to 5 or in claim 6 or claim 7 when dependent directly or indirectly upon any one of claims 3 to 5.

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FIG. 1

5' GGGGGGGGGGGGGGG CA CCA CCA CTG ACC
10 20

-24 -20 -10
Met Pro Ser Ser Val Ser Trp Gly Ile Leu Leu Leu Ala-Gly Leu
TGG GAC ACT GAA TCC ACA ATG CGG TCT TCT GTC TCG TGG GGC ATC CTC CTG CTG GCA GGC CTG
30 40 50 60 70 80 90

-1 +1 10
Cys Cys Leu Val Pro Val Ser Leu Ala Glu Asp-Pro Gln Gly Asp Ala Ala Gln Lys Thr Asp
TGC TGC CTG GTC CCT GTC TCC CTG GCT GAG GAT CCC CAG GGA GAT GCT GCC CAG AAC ACA GAT
100 110 120 130 140 150

20 30
Thr Ser His His Asp Gln Asp His Pro Thr Phe Asn Lys Ile Thr Pro Asn Leu Ala Glu Phe
ACA TCC CAC CAT GAT CAG GAT CAC CCA ACC TTC AAC AAG ATC ACC CCC AAC CTG GCT GAG TTC
160 170 180 190 200 210

40 50
Ala Phe Ser Leu Tyr Arg Gln Leu Ala His Gln Ser Asn Ser Thr Asn Ile Phe Phe Ser Pro
GCC TTC AGC CTC TAC CGC CAG CTG GCA CAC CAG TCC AAC ACC ACC AAT ATC TTC TTC TCC CCA
220 230 240 250 260 270 280

60 70
Val Ser Ile Ala Thr Ala Phe Ala Met Leu Ser Leu Gly Thr Lys Ala Asp Thr His Asp Glu
GTG AGC ATC GCT ACA GCC TTT GCA ATG CTC TCC CTG GGG ACC AAG GCT GAC ACT CAC GAT GAA
290 300 310 320 330 340

80 90
Ile Leu Glu Gly Leu Asn Phe Asn Leu Thr Glu Ile Pro Glu Ala Gln Ile His Glu Gly Phe
ATC CTG GAG GCC CTG AAT TTC AAC CTC ACG GAG ATT CCG GAG GCT CAG ATC CAT GAA GGC TTC
350 360 370 380 390 400

100 110
Gln Glu Leu Leu Arg Thr Leu Asn Gln Pro Asp Ser Gln Leu Gln Leu Thr Thr Gly Asn Gly
CAG GAA CTC CTC CGT ACC CTC AAC CAG CCA GAC AGC CAG CTC CAG CTG ACC ACC GGC AAT GGC
410 420 430 440 450 460 470

120 130
Leu Phe Leu Ser Glu Gly Leu Lys Leu Val Asp Lys Phe Leu Glu Asp Val Lys Lys Leu Tyr
CTG TTC CTC ACC GAG GCC CTG AAG CTC GTG GAT AAG TTT TTG GAG GAT GTT AAA AAG TTG TAC
480 490 500 510 520 530

140 150 160 170 180
His Ser Glu Ala Phe Thr Val Asn Phe Gly Asp Thr Glu Glu Ala Lys Lys Gln Ile Asn Asp
CAC TCA GAA GCC TTC ACT GTC AAC TTC GGG GAC ACC GAA GAG GCC AAG AAA CAG ATC AAC GAT
540 550 560 570 580 590

160 170 180
Tyr Val Glu Lys Gly Thr Gln Gly Lys Ile Val Asp Leu Val Lys Glu Leu Asp Arg Asp Thr
TAC GTG GAG AAG GGT ACT CAA GGC AAA ATT GTG GAT TTG GTC AAG GAG CTT GAC AGA GAC ACA
600 610 620 630 640 650

190 200
Val-Phe Ala Leu Val Asn Tyr Ile Phe Phe Lys Gly Lys Trp Glu Arg Pro Phe Glu Val Lys
GTT TTT GCT CTG GTG AAT TAC ATC TTC TTT AAA GCC AAA TCG GAG AGA CCC TTT GAA GTC AAG
660 670 680 690 700 710 720

210 220
Asp Thr Glu Glu Glu Asp Phe His Val Asp Gln Val Thr Val Lys Val Pro Met Met Lys
GAC ACC GAG GAA GAG GAC TTC CAC GTG GAC CAG GTG ACC ACC GTG AAG GTG CCT ATG AIG AAC
730 740 750 760 770 780

230 240
Arg Leu Gly Met Phe Asn Ile Gln His Cys Lys Lys Leu Ser Ser Trp Val Leu Leu Met Lys
CGT TTA GGC ATG TTT AAC ATC CAG CAC TCT AAG AAC CTC AGC TGG GTG CTG CTG CTG AIG AAA
790 800 810 820 830 840

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FIG. I - page 2

Tyr Leu Gly Asn Ala Thr Ala Ile Phe Phe Leu Pro Asp Glu Gly Lys Leu Gln His Leu Glu
 TAC CTG GGC AAT GCC ACC GCC ATC TTC TIC CTG CCT GAT GAG GGG AAA CTA CAG CAC CTG GAA
 850 860 870 880 890 900 910
 Asn Glu Leu Thr His Asp Ile Ile Thr Lys Phe Leu Glu Asn Glu Asp Arg Arg Ser Ala Ser
 AAT GAA CTC ACC CAC GAT ATC ATC ACC AAC TTC CTG GAA AAT GAA GAC AGA AGG TCT GCC AGC
 920 930 940 950 960 970
 Leu His Leu Pro Lys Leu Ser Ile Thr Gly Thr Tyr Asp Leu Lys Ser Val Leu Gly Gln Leu
 TTA CAT TTA CCC AAA CTG TCC ATT ACT GGA ACC TAT GAT CTG AAG AGC GTC CTG GGT CAA CTG
 980 990 1000 1010 1020 1030
 Gly Ile Thr Lys Val Phe Ser Asn Gly Ala Asp Leu Ser Gly Val Thr Glu Glu Ala Pro Leu
 GCC ATC ACT AAG GTC TTC AGC AAT GGG GCT GAC CTC TCC GGG GTC ACA GAG GAG GCA CCC CTG
 1040 1050 1060 1070 1080 1090 1100
 Lys Leu Ser Lys Ala Val His Lys Ala Val Leu Thr Ile Asp Glu Lys Gly Thr Glu Ala Ala
 AAC CTC TCC AAG GCC GTG CAT AAG GCT GTG CTG ACC ATC GAC GAG AAA GGG ACT GAA GCT GCT
 1110 1120 1130 1140 1150 1160 1170
 Gly Ala Met Phe Leu Glu Ala Ile Pro Met Ser Ile Pro Pro Glu Val Lys Phe Asn Lys Pro
 GGG GCC ATC TTT TTA GAG GCC ATA CCC ATG TCT ATC CCC CCC GAG GTC AAG TTC AAC AAA CCC
 1170 1180 1190 1200 1210 1220 1230
 Phe Val Phe Leu Met Ile Glu Gln Asn Thr Lys Ser Pro Leu Phe Met Gly Lys Val Val Asn
 TTT GTC TTC TTA ATG ATT GAA CAA AAT ACC AAG TCT CCC CTC TIC ATG GGA AAA GIG GIG AAT
 1230 1240 1250 1260 1270 1280 1290
 Pro Thr Gln Lys STOP
 CCC ACC CAA AAA TAA CTG CCT CTC GCT CCT CAA CCC CTC CCC TCC ATC CCT GGC CCC CTC CCT
 1290 1300 1310 1320 1330 1340 1350
 GGA TGA CAT TAA AGA AGG GTT GAG CTG
 1360 1370

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ATG ACC ATG ATT ACG CCA AGC TTG GGC TGC AGG TCG ACT CTA GAG GAT CCC CGG GCG AGG TCG AAT TCA CTG GCC
HindIII PstI SalI XbaI BamHI XbaI SstI EcoRI HaeIII SmaI AccI, HincII

1 2 3 4 PRO SER LEU GLY CYS ARG SER THR LEU GLU ASP PRO ARG ALA SER SER 5 6 7 8

THR MET ILE THR ASN SER LEU ALA

ATG ACC

ATG ACC ATG ATT ACG CCA AGC TTG GGC TGC AGG TCG ACT CTA GAG GAT CCC CGG GCG AGG TCG AAT TCA CTG GCC

HindIII

PstI

SalI

XbaI

BamHI

XbaI

SstI

EcoRI

HaeIII

SmaI

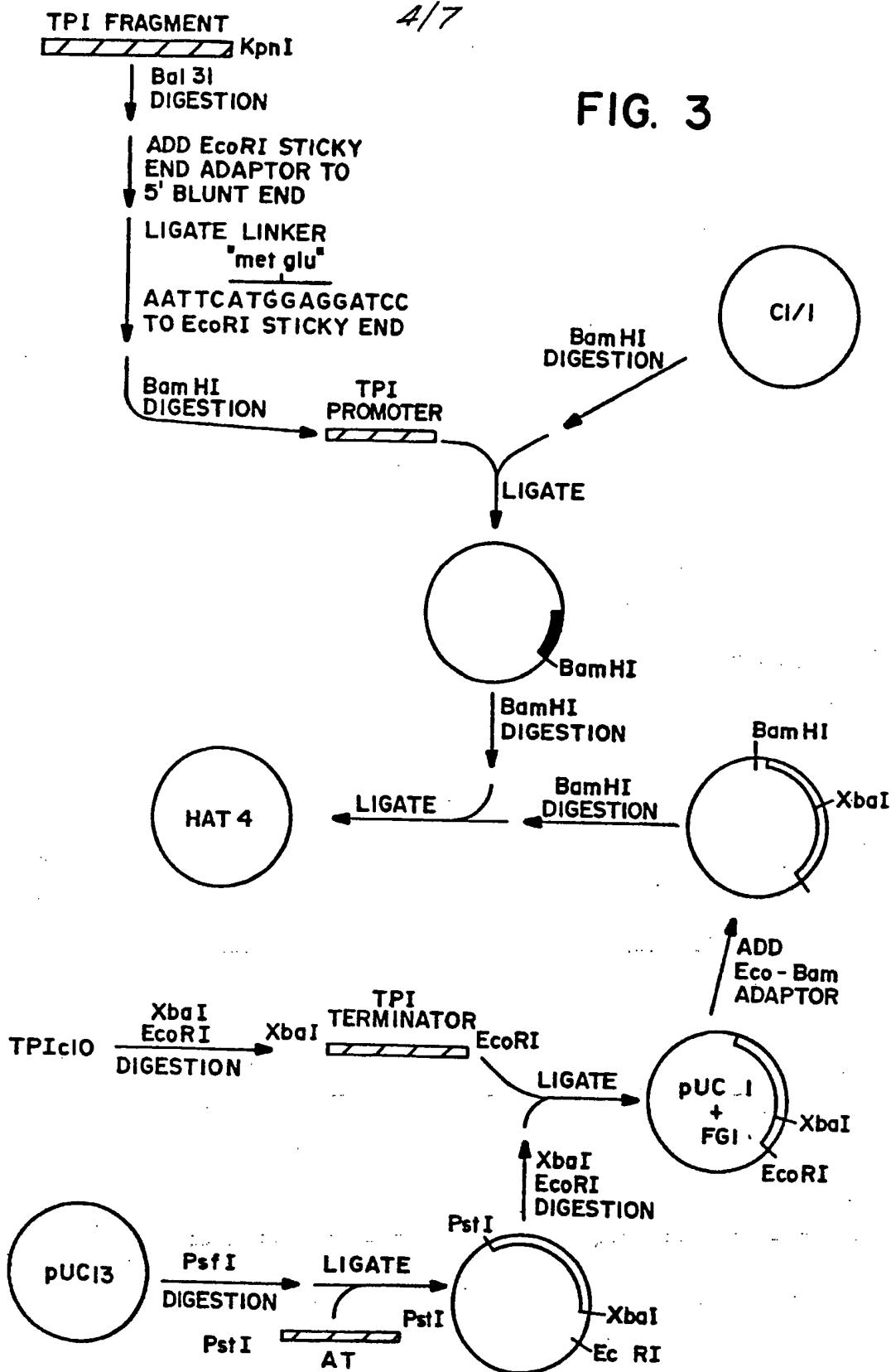
AccI, HincII

FIG. 2

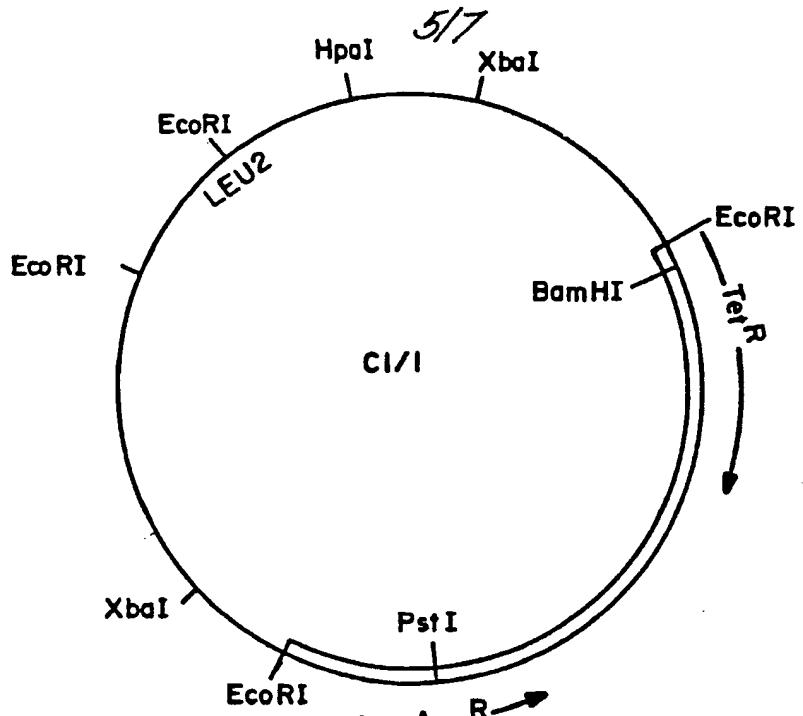
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FIG. 3



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■ pBR322

FIG. 4

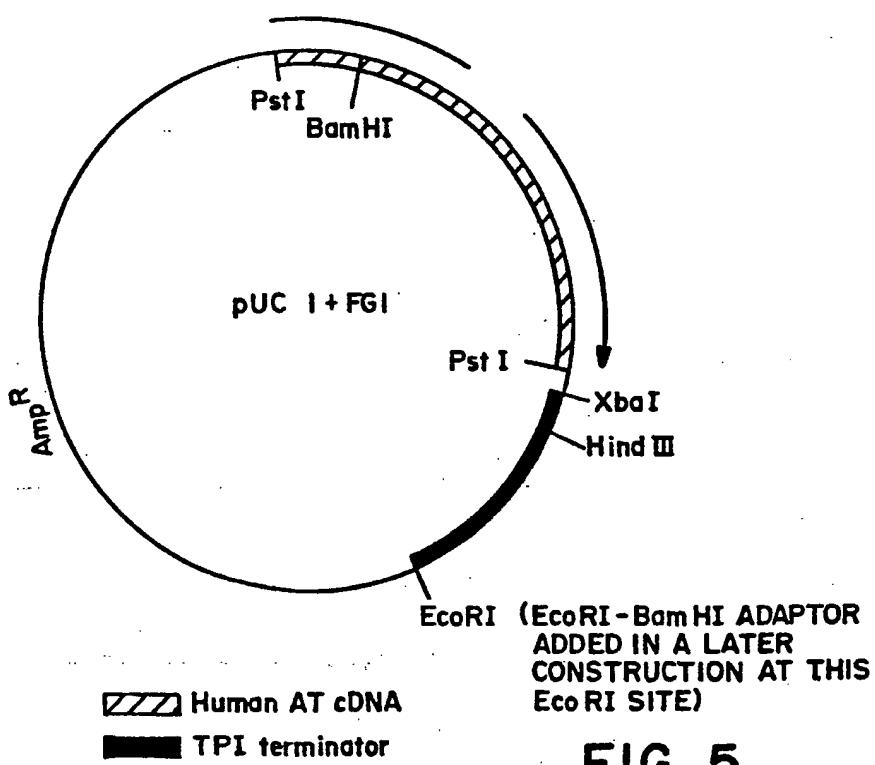


FIG. 5

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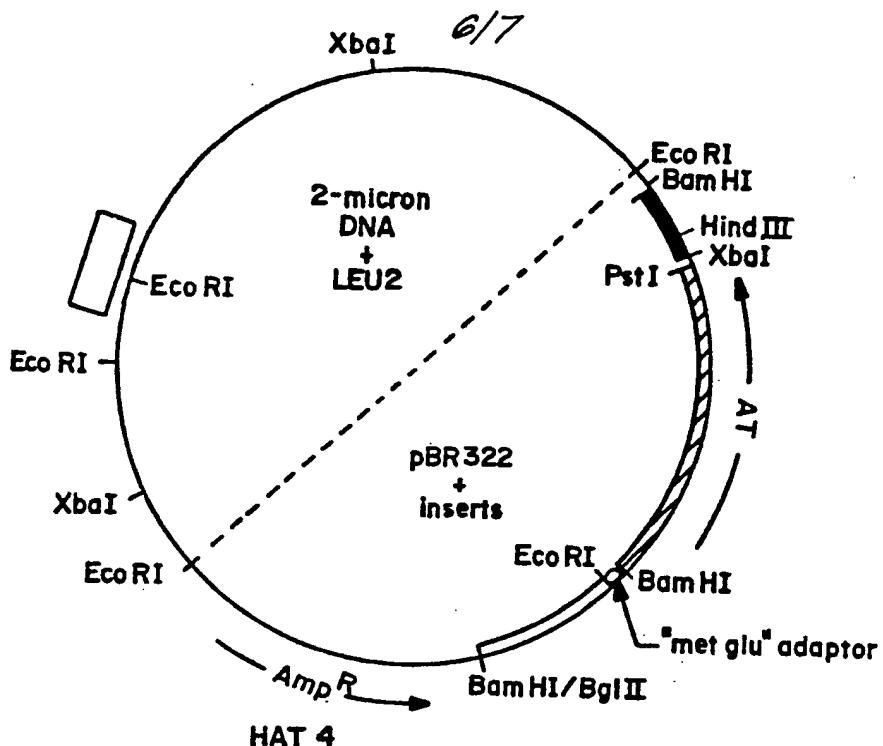


FIG. 6

- TPI promoter
- ▨ Human AT cDNA
- TPI terminator

PR 103-5

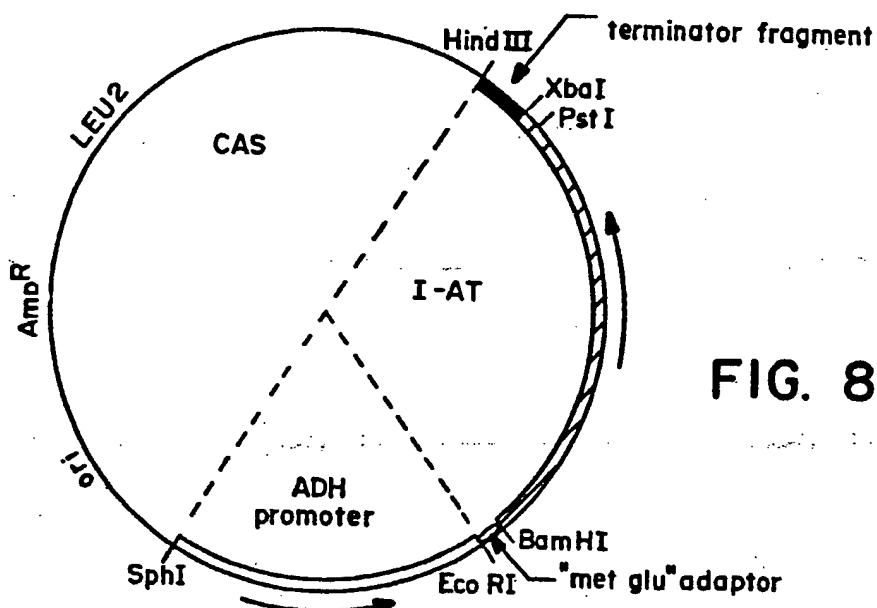


FIG. 8

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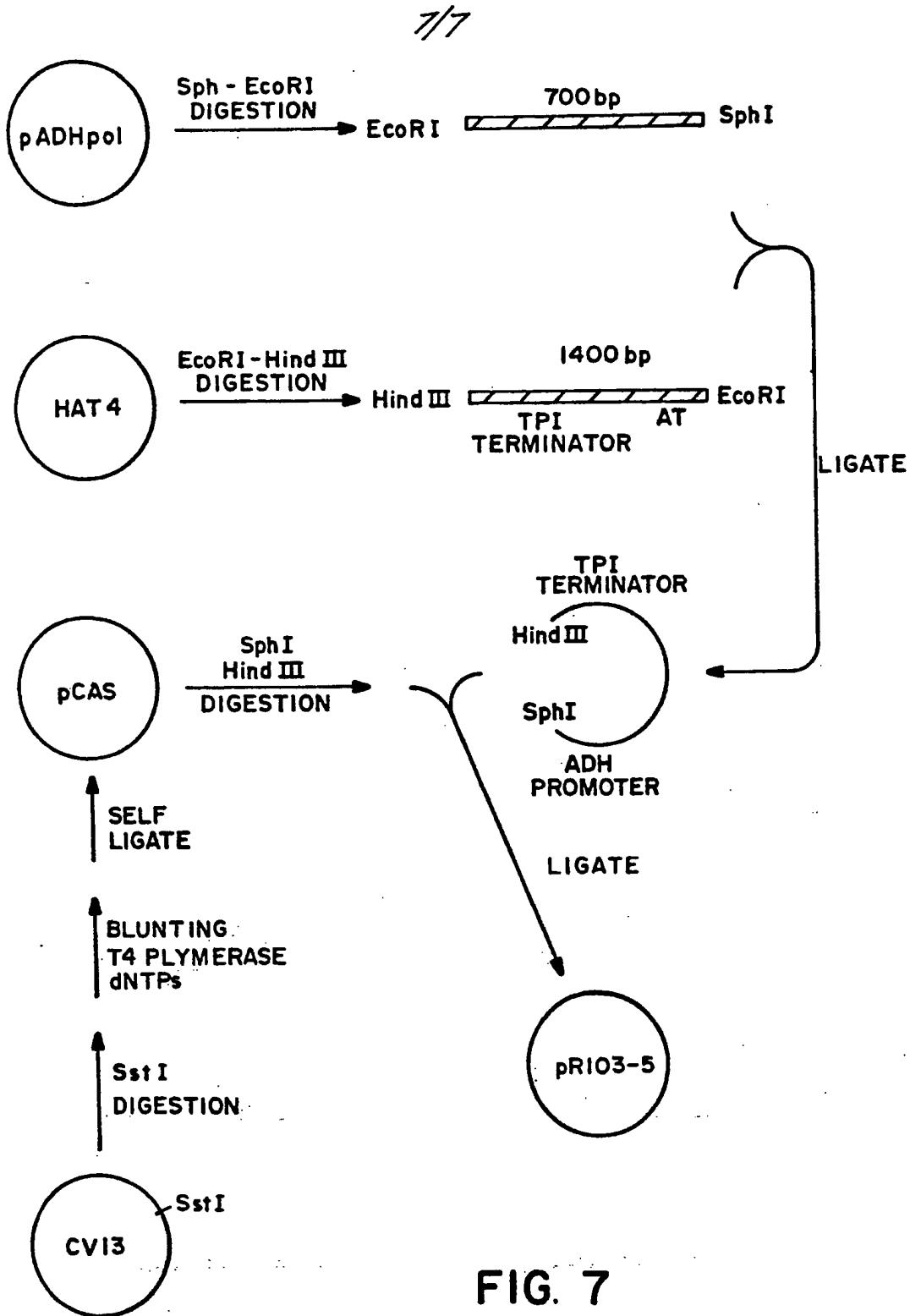


FIG. 7



European Patent
Office

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Application number

EUROPEAN SEARCH REPORT

DOCUMENTS CONSIDERED TO BE RELEVANT			EP 84305571.6
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
A	<p>NATURE, vol. 290, no. 5802, March 12, 1981 (New York, London)</p> <p>D. BEACH et al. "High-frequency transformation of the fission yeast <i>Schizosaccharomyces pombe</i>" pages 140-142</p> <p>* Totality *</p> <p>--</p>	1,8, 11,13	<p>C 12 N 15/00</p> <p>C 12 P 21/00</p> <p>C 12 N 9/99</p> <p>A 61 K 37/02</p> <p>//C 12 R 1/85</p>
	<p>MOLECULAR & GENERAL GENETICS, vol. 182, no. 3, 1981 (Springer Verlag, Berlin, Heidelberg)</p> <p>M. YAMAMOTO et al. "Cloning of a Gene from the Fission Yeast <i>S. pombe</i> which Complements <i>E. coli</i> pyr B, the Gene for Aspartate Transcarbamylase" pages 426-429</p> <p>* Totality *</p> <p>--</p>		
D,A	<p>PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, vol. 78, no. 11, November 1981 (Baltimore, USA)</p> <p>K. KURACHI et al. "Cloning and sequence of cDNA coding for α_1-antitrypsin" pages 6826-6830</p> <p>* Abstract *</p> <p>--</p>	1,3,4, 11,13	<p>TECHNICAL FIELDS SEARCHED (Int. Cl.4)</p> <p>C 12 N</p> <p>C 12 P</p> <p>A 61 K</p> <p>C 07 H</p>
	<p>The present search report has been drawn up for all claims</p>		
	<p>Place of search VIENNA</p>		<p>Date of completion of the search 23-11-1984</p>
CATEGORY OF CITED DOCUMENTS		<p>T : theory or principle underlying the invention</p> <p>E : earlier patent document, but published on, or after the filing date</p> <p>D : document cited in the application</p> <p>L : document cited for other reasons</p> <p>R : member of the same patent family, corresponding document</p>	
<p>X : particularly relevant if taken alone</p> <p>Y : particularly relevant if combined with another document of the same category</p> <p>A : technological background</p> <p>O : non-written disclosure</p> <p>P : intermediate document</p>			



European Patent
Office

EUROPEAN SEARCH REPORT

0139383

Application number

DOCUMENTS CONSIDERED TO BE RELEVANT			EP 84305571.6
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
D, A	<p>BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 103, no. 2, November 30, 1981 (New York, London, Toronto, Sydney, San Francisco, Tokyo)</p> <p>T. CHANDRA et al. "Induction of α_1-Antitrypsin mRNA and cloning of its cDNA" pages 751-758</p> <p>* Summary *</p> <p>-----</p>	1, 3, 4, 11, 13	
			TECHNICAL FIELDS SEARCHED (Int. Cl.4)
<p>The present search report has been drawn up for all claims</p>			
Place of search	Date of completion of the search	Examiner	
VIENNA	23-11-1984	WOLF	
CATEGORY OF CITED DOCUMENTS		<p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document</p>	
<p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p>			